

# Novel 3-D Sample Plate using Monolithic Capture Media in Collimated-Hole Structures for Interfacing High Capacity Separations with MALDI-TOF

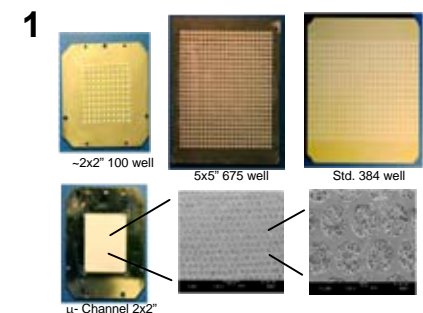
Poster Number TP 060

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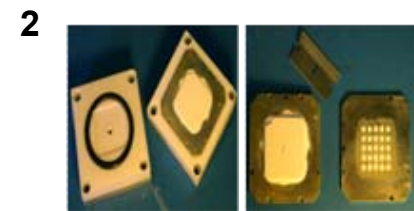
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## Overview/Introduction:

- Novel MALDI targets are being developed that use collimated hole structures (CHS) combined with monolithic chromatography media to enable capture & concentration of sample and serve as a direct interface between the mass spectrometer and different separation schemes -HPLC, electrophoresis and tissue imaging-
- Plate construction, sample deposition (LC) and sample elution are presented
- Results of capacity and LC interface at 15 and 50  $\mu\text{L}/\text{min}$  separation speeds are presented



**Figure 1: CHS plates:** Row 1: 100, 675 and 384 wells plate of 1.5, 3 and 10 mm thickness respectively constructed in metal and plastic designed for modes of discrete sample deposition such as LC or robotic and hand spotting. Row two shows glass  $\mu$ -channel CHS plate (25  $\mu\text{m}$  holes) designed for interface with gel and tissue applications.



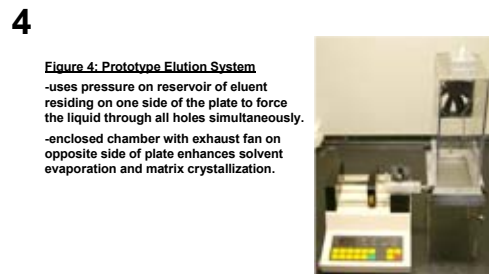
## Figure 2: Plate Construction:

- styrene/divinylbenzene based polymers for reversed phase capture media of protein/peptides - monomer solutions are injected into tetrafluoroethylene molds
- polymerization is thermally initiated
- Excess polymer is shaved to a level coincident with the plate substrate (metal, plastic, glass). -ratio and constitution of dilution solvent(porogens) are used to tailor pore-size/flow properties -Sample application/analysis occur on the polymer surface.



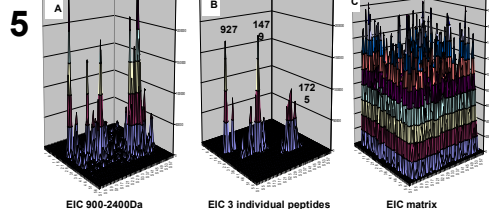
**Figure 3: Prototype LC deposition System**

- X Y control of solvent capillary
- Tip seals with plate surface
- column pressure forces column effluent through discrete locations on CHS plates.



**Figure 4: Prototype Elution System**

- uses pressure on reservoir of eluent residing on one side of the plate to force the liquid through all holes simultaneously.
- enclosed chamber with exhaust fan on opposite side of plate enhances solvent evaporation and matrix crystallization.

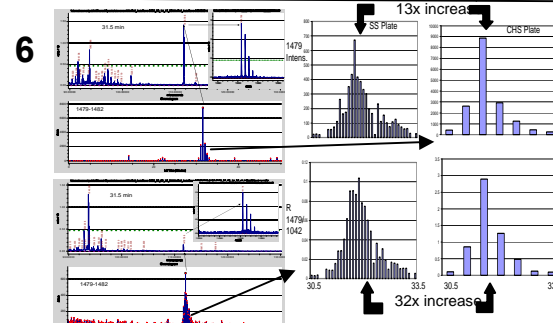


**Figure 5 LC Results:**

- 3D plots of signal intensity as function of surface location from 1 pmol BSA digest LC separation Conditions: 0.5 mm column ID C-18 (eksigent), 15  $\mu\text{L}/\text{min}$ , 5-45%ACN gradient (50min), dwell time 30s/spot 10x10 CHS plate (Fig. 1 top left)

- Frame A: extracted ion chromatogram (EIC) all peptides from 900-2400 Da.
- Frame B: EIC of three individual peptides (927,1479,1725 Da)
- Frame C: EIC of 379Da ACCH matrix dimer.

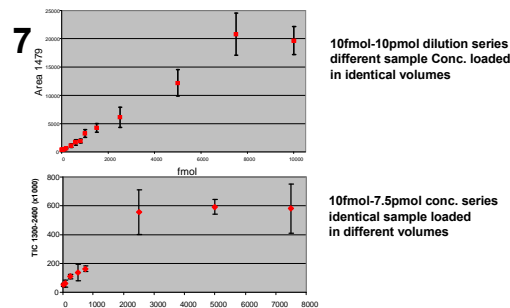
Figure demonstrates the ability of the CHS plates to capture/concentrate sample without loss of chromatographic resolution and uniform sample elution.



**Figure 6: Comparison with conventional 2D plate:**

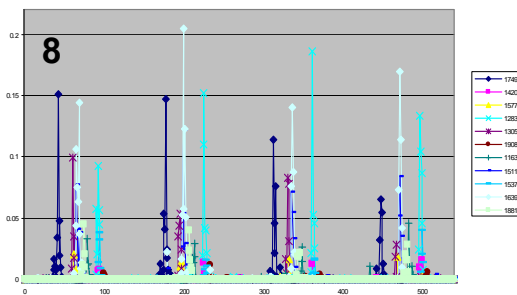
- Identical 1hr separation of 1 pmol BSA at 15  $\mu\text{L}/\text{min}$  flow spotted at 5s intervals on a 2D surface and 20s intervals on a CHS plate.
- Left frames shows the EIC and max-intensity spectrum of 1479Da peptide from each run (CHS top;2D bottom).
- Right frames are blow-ups of the EIC region of interest.
- Right top: raw signal intensity 13x increase in intensity
- Right bottom: signal intensity normalized against a 1042Da internal standard (matrix addition)
- demonstrates 32x increase in signal intensity

Based dwell time and spot dimensions a 36x increase in signal was calculated therefore the above results indicate near 100% sample recovery.



**Figure 7: Capacity study:**

- plots from different loading schemes aimed at determining capacity of CHS plate with  $\sim 5 \mu\text{L}$  void volume.
- Plot A: dilution series of different conc. loaded in some volume as not overwhelm the void volume of well shows  $\sim 8$  pmol load capacity limited primarily by analyte/matrix ratio/ ionization efficiency
- Plot B Same sample different load volume  $\sim$ void volume exceeded-- Limited by binding capacity of CHS wells  $\sim 2.5$  pmol of digested protein.
- Binding capacity will vary with well dimensions



**Figure 8: Four identical 25 min, 50  $\mu\text{L}/\text{min}$  LC separation on single CHS plate**  
Conditions: 30x1mm, 200Å, C-18 5  $\mu\text{m}$ , Higgins Anal., 0-45% ACN (25 min) 50  $\mu\text{L}/\text{min}$  flow, dwell time/spot 10s  
-overlay of EIC from 12 mid-intensity range peptides plotted as ratio against 1042 internal standard and show average relative standard deviation of 22%.  
-5 pmol of BSA digest run in series and spotted on a single 675 well CHS plate (Fig. 1 top middle)  
-separations conditions represent a 50-100X increase in flow rate/sample load and capture over conventional LC-MALDI schemes performed on 2D plates.  
-relative large sample load, high speed, reproducible separation afforded by CHS plates.

## Conclusions:

- Novel 3-dimensional MALDI target that captures and concentrates peptide/protein sample and serves as a direct interface between separation and mass spectrometer
- Plates allow for higher load capacity (potentially 100s  $\mu\text{g}$  to mg quantity) and higher flow (100s  $\mu\text{L} - \text{mL}/\text{min}$ ) separations that should enable dramatic increase in the detection levels of lower concentration proteins/peptides in complex proteomic mixtures.
- Plates are designed to be reusable with current prototypes easily withstanding 25-50 analyses (like typical LC columns).
- Presentation focuses on the reversed phase LC-MALDI interface because of its near ubiquitous capture of peptide material; however, plates focused on affinity capture (ie. glyco-capture) and studies focused on direct interface of Gel and tissue sample preparation schemes with MALDI are underway.

## Acknowledgement:

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